

REVIEW

Open Access



# Immunostimulants in respiratory diseases: focus on Pidotimod

Francesca Puggioni<sup>1,2\*</sup>, Magna Alves-Correia<sup>2,3</sup>, Manar-Farouk Mohamed<sup>2,4</sup>, Niccolò Stomeo<sup>1</sup>, Riccardo Mager<sup>1</sup>, Massimiliano Marinoni<sup>1</sup>, Francesca Racca<sup>2</sup>, Giovanni Paoletti<sup>1,2</sup>, Gilda Varricchi<sup>5</sup>, Veronica Giorgis<sup>2</sup>, Giovanni Melioli<sup>1,2</sup>, Giorgio Walter Canonica<sup>1,2</sup> and Enrico Heffler<sup>1,2</sup>

## Abstract

Usefulness of Pidotimod and its role as immunostimulant, has been discussed, we know, for several decades. Nevertheless, there is still much to know. Understanding its mechanisms and its potential usefulness in airway infections and its prevention, asthma both Th2 and non Th2 type, bronchiectasis, as adjuvant in vaccination and in allergen immunotherapy still remains to clearly unveil. The aim of this paper was to provide a useful updated review of the role of the main available immunostimulants, giving particular focus on Pidotimod use and its potentials utility in respiratory diseases. Pidotimod showed its usefulness in reducing need for antibiotics in airway infections, increasing the level of immunoglobulins (IgA, IgM, IgG) and T-lymphocyte subsets (CD3+, CD4+) endowed with immunomodulatory activity that affect both innate and adaptive immune responses. Higher expression of TLR2 and of HLA-DR molecules, induction of dendritic cell maturation and release of pro-inflammatory molecules, stimulation of T lymphocyte proliferation and differentiation toward a Th1 phenotype, as well as an increase of the phagocytosis have been demonstrated to be associated with Pidotimod in *in vitro* studies. All these activities are potentially useful for several respiratory conditions such as asthma, COPD, and recurrent respiratory tract infections.

**Keywords:** Pidotimod, Immunostimulants, Respiratory diseases, Adaptive immunity, Innate immunity, Allergy, Asthma, Chronic obstructive lung disease

## Introduction

Since the early 90's more than a hundred papers have been published about Pidotimod, an immunostimulant drug. Due to its action mechanism and its potential, it has been used both in children and in adults, mostly for preventing respiratory tract infections, and asthma or chronic obstructive pulmonary disease exacerbations.

Despite the use of antibiotics and vaccines, the frequency of respiratory tract infections is still high and these infections disturb a wide range of patients, from children to elderly, particularly these two extremes due to the deficiency of their immune system: immaturity in the first case and "immunosenescence" in the second one. For that reason, immunostimulant drugs steadily

increased in the past few years, getting nowadays more importance and visibility in preventing the onset and reducing the duration of airway infections [1].

Immunostimulants are a heterogeneous group of compounds that act non-specifically on the immune system by inducing its activation, either upregulating it or by favoring the activity of one of its components. By understanding deeply their biological function, they may be implemented in the clinical practice to shape the immune system favorably according to the different processes that want to be enhanced or hampered. According to these peculiar characteristics they have also been called "Biological Response Modifiers" (BRMs). Among this heterogeneous group of compounds which encompass both synthetic and naturally occurring substances, some of them deserve a brief discussion [2–31]. This review article will briefly summarize the most commonly used BRMs used in the treatment of respiratory diseases, and it will focus mainly on pidotimod and its current and potential clinical use.

\* Correspondence: [francesca.puggioni@humanitas.it](mailto:francesca.puggioni@humanitas.it)

<sup>1</sup>Department of Biomedical Sciences, Humanitas University, Pieve Emanuele, MI, Italy

<sup>2</sup>Personalized Medicine, Allergy and Asthma - Humanitas Clinical and Research Center – IRCCS, Via Alessandro Manzoni 56, 20089 Rozzano, MI, Italy  
Full list of author information is available at the end of the article



*Bacterial Lysates* derive from a blend of bacterial cultures antigens throughout mechanical or chemical cell lysis. The selected bacteria include: *Streptococcus pneumoniae*, *Streptococcus pyogenes*, *Hemophilus influenzae*, *Legionella pneumophila*, *Moraxella catarrhalis*, *Mycoplasma pneumoniae*, *Chlamydia pneumoniae*, *Streptococcus Aureus*, *Streptococcus viridans* and *Klebsiella pneumoniae*. These bacterial antigens have shown to directly upregulate both innate and adaptive immunity, by stimulating Toll-like receptors, inducing dendritic cells (DC) maturation, increasing phagocytic activity, and by activating interleukin 2 receptor (IL2R) on different lymphocytes subsets, inducing therefore cytokines synthesis and enhancing the production of IgA antibodies [2–4].

A Cochrane review by Del-Rio-Navarro et al. concluded that immunostimulants reduce the incidence of acute respiratory tract infections (ARTIs) by 40% in susceptible children, but the included trials quality was generally poor with a high level of statistical heterogeneity [5]; however, the subgroup analysis of bacterial lysates studies showed lower heterogeneity and displayed slightly better quality [5] and this was confirmed by a subsequent systematic review on OM-85 BV, a bacterial lysate [6]. Other beneficial effects of bacterial lysate in children are: reduction in the usage of antibiotics and duration of infectious episodes [7], faster improvement of the symptoms, shorter convalescence and decreased incidence of consequent infectious episodes when administered in children with sub-acute sinusitis [8].

In adults and elderly, polyvalent mechanical bacterial lysates (PMBL) have been shown to be superior respect to placebo and to polyvalent chemical bacterial lysates (PCML) in terms of reducing the number and the duration of infectious episodes, and reducing the need of antibiotic treatment in patients with recurrent respiratory tract infections (RRTI) [9]. Moreover, the high pressures used in PMBL production can eliminate some chemical contaminants with a lower degree of damage to bacterial antigens. PMBLs have been also shown to be beneficial in chronic obstructive pulmonary disease (COPD) patients by reducing the days of hospitalizations, increasing the interval between two exacerbations and reducing the number of days with fever; nonetheless they failed to reduce the number of exacerbations by 25% [10].

*Lactoferrin* is a natural iron-binding protein highly expressed by both epithelial cells in most exocrine secretions, and by neutrophils, which store the molecule in the secondary granules. It has been reported its functions as an alarmin-like molecule, promoting antigen presenting cells maturation and upregulation of pro-inflammatory cytokines synthesis [11], but also as a potent anti-inflammatory molecule on monocytes by triggering

tolerogenic-like program during their differentiation into DC [12]; other reported functions of lactoferrin are: an inhibitory effect on eosinophils migration [13], interference with mast cells functions [14], and an effect on the immune Th1/Th2 balance [15].

The clinical effect of lactoferrin supplementation as a preventive measure for infectious diseases has been mainly investigated in newborn and preterm infants. Notably, an Italian study conducted by Manzoni et al. showed a significant decrease in the incidence of sepsis in very low birth weight infants (< 1500 g) who received 100 mg/d of bovine lactoferrin from birth until the 30 or 45 day of life [16].

*Resveratrol* is a natural polyphenolic compound contained in several plant species, such as peanuts, grapes and berries. It has been shown that resveratrol has an effect on a wide range of biological activities ranging from antimicrobial [17], neuroprotection [18] and anti-inflammatory [19]. Resveratrol's molecular interactions still remain largely unknown, however it has been shown to act as an activator of sirtuin-1 (which is thought to play a role in activating Th17 cells) [20], to modulate T-Reg/Th17 balance [21] and to downregulate pro-inflammatory cytokines production [22].

Notably, it has been demonstrated that resveratrol has a remarkable anti-inflammatory effect in COPD patients and its antimicrobial and immunomodulatory effects have been studied *in vivo* with a septicemia infection model system induced by nontypeable *Haemophilus influenzae* (NTHi), showing a reduction in NTHi viability and a concomitant reduction in airway inflammation [23]. Furthermore, in a recent study conducted by Wu S et al, resveratrol reduced mortality, lung injury and cytokine levels in the lung tissue of *S. aureus* pneumonia in murine models [24]. Based on these studies, Resveratrol may be useful in the acute phase of pulmonary infections.

*Monophosphoryl lipid A* (MPLA) is a nontoxic, but still highly immunogenic derivative of lipid A, the biologically active part of lipopolysaccharide (LPS) endotoxin, and a Toll-like receptor 4 agonist. *In vitro* studies confirmed that MPLA is able to act, as its parent molecule LPS, as a potent adjuvant increasing the function of antigen presenting cells and an antibody response characterized by the production of IgG1 and IgG2 [25]. Because of its immunostimulant properties, MPLA has been used as adjuvant in malaria vaccine preparations [26], in hepatitis B surface antigen vaccine [27] and in papilloma virus vaccine [28]. Furthermore, it has been shown that MPLA promote allergen-induced immune deviation in favor of Th1 responses, confirming its beneficial role as an adjuvant in allergen immunotherapy [29, 30]. Interestingly, beside its role as adjuvant molecule, it has been demonstrated that MPLA administration

before or after the induction of systemic bacterial infection results in an improved survival and an increased bacterial clearance [31, 32].

### **Pidotimod**

Pidotimod is a synthetic dipeptide molecule (3-l-pyrroglutamyl-l-thiazolidine-4-carboxylic acid) endowed with immunomodulatory activity that affects both innate and adaptive immune responses. Higher expression of Toll Like Receptors (TLR) 2 and of HLA-DR molecules, induction of DC maturation and release of pro-inflammatory molecules, stimulation of T lymphocyte proliferation and differentiation toward a Th1 phenotype, as well as an increase in the phagocytosis have been demonstrated to be associated with Pidotimod in *in vitro* studies [1, 33–35].

Several *in vitro* and *in vivo* studies on human and animals showed that pidotimod is able to ameliorate both innate and adaptive immunity, and enhances the immune system capabilities to fight RRTI, especially in children with increasing resistance to viral infections [36–40].

Studies in different areas have demonstrated the benefit of Pidotimod, including its use in hepatitis C, HPV genital infection, Henoch-Schönlein Purpura, nephrotic syndrome, and immunodepressed individuals such as children and elderly [1]. Considering its good safety profile, only one single case of potentially severe adverse event (new onset of Henoch-Schönlein Purpura) was reported to be associated to the intake of Pidotimod [41].

The common end-point of these studies is that Pidotimod has an immunomodulatory activity which is able both to improve the clinical conditions of patients and to enhance and stimulate their immunity cells functions acting on both adaptive and innate immunity [1].

The clinical efficacy of pidotimod in decreasing respiratory tract infections in paediatric age was assessed in multicenter randomized trials involving children with age ranged from 2 to 14 years old as regard fever duration, need to antibiotic treatment, hospitalization, school absence, recovery time and relapse rate. All these parameters were significantly reduced in the Pidotimod-treated groups, clarifying the supportive role of pidotimod in treatment of acute infections and its prophylactic role in prevention of RRTI [42, 43].

### **Effect of pidotimod on innate immunity**

It is well known that epithelial cells play a primary defensive role, not only providing a natural physical barrier but also being involved in innate and adaptive immune responses [1], by means of several mechanisms including the expression of TLRs on their surface. TLRs are pattern recognition receptors of molecules that are broadly shared by pathogens: these molecules are called pathogen-associated molecular patterns (PAMPs) [44, 45].

Recognition of PAMPs via TLRs results in the activation of NF- $\kappa$ B, which is a complex protein found in almost all animal cells and controls DNA transcription in response to variable stimuli, including bacterial and viral infections [46, 47]. Activated NF- $\kappa$ B leads to stimulation of innate immune cells to release different cytokines and chemokines, eventually leading to internalization and phagocytosis of the attached pathogens [44, 45].

Carta et al designed an *in vitro* study to evaluate the immunomodulatory effects of pidotimod demonstrating that it is able to upregulate TLR2 expression, with significant increase in NF- $\kappa$ B protein expression and NF- $\kappa$ B nuclear translocation [48].

Pidotimod is also able to induce phenotypic and functional maturation of mucosal DC, which plays an important role in the cross talk between innate and adaptive immunity, as pidotimod upregulates the expression of HLA-DR and co-stimulatory surface markers CD83, CD88.

Pidotimod-induced DC maturation leads to release of pro-inflammatory mediators such as TNF- $\alpha$  leading to increase recruitment of inflammatory cells, activation of naïve T lymphocytes with proliferation and polarization toward Th1 phenotype. Besides its role on epithelial cells and DC, pidotimod also enhances cytotoxic activity of natural killer cells (NK) and phagocytic activity of neutrophils [49].

### **Effect of Pidotimod on adaptive immunity**

Recent immunological data concluded that functional disorder of Th1/Th2 cells could be strictly related to development of RRTI in children. Zhou and Dai proposed a study comparing the effect of Pidotimod to spleen aminopeptide (a cell immunity enhancer made of peptides and nucleotides extracted from spleen of healthy animals) on Th1/Th2 cytokines balance in children with RRTI [50]. Pidotimod was significantly associated with enhancement of secretion of IFN- $\gamma$  and other Th1 cytokines such as IL12 through which Th1 mediates its inflammatory reactions and delayed hypersensitivity reactions more than spleen aminopeptide, on the same time it was able to down regulate IL4 secretion which is essential for Th2 activity [50].

It is appropriate to mention that Pidotimod effects on decreasing Th2 cytokines against increasing Th1 cytokines would not only enhance the immune system capabilities to fight infections, but it has also a protective role against the development of atopy; pidotimod also was found to down regulate CD30 expression on cells which is linked to Th2 cells [51].

A cohort randomized study held on children with Down syndrome (who are more prone to develop frequent respiratory tract infections), demonstrated that Pidotimod significantly upregulates genes involved in

inflammation, chemotaxis and antimicrobial activity. Moreover, the ratio of flu-specific immunoglobulin G1/G3 (IgG1/IgG3) was skewed in Pidotimod treated individuals when it was given in association to seasonal virosomal adjuvanted influenza vaccine [52].

Moreover, increased production of nasopharyngeal and salivary secretory IgA (sIgA) in children with respiratory tract infections treated with Pidotimod has been demonstrated, letting suppose a possible direct effect of pidotimod on B lymphocytes [53].

Another study used nasal cytology to evaluate the ability of pidotimod to reduce nasal inflammation and improve children quality of life through a randomized controlled study involved 40 children with RRTI. Nasal cell motility, mucociliary clearance and the presence of supra nuclear-stria (SNS) were used as an indirect index of wellness of nasal ciliated cells: an improvement was seen in all these cytological features in the pidotimod treated group, with concomitant decreased frequency of respiratory tract infections and antibiotics consumption [54].

The main mechanisms of action of pidotimod, at both innate and adaptive immunity levels, are summarized in Fig. 1.

#### Pharmacodynamics and safety of pidotimod

Pidotimod follows a first order pharmacokinetic when administrated through oral route, while parenteral route follows second order pharmacokinetics, with a half-life of 4 h with and an oral bio-availability of 42–44%. It is eliminated through the kidneys without being metabolized. Plasma clearance is  $51 \times \text{hE-1}$  with apparent distribution volume 30l. Several types of oral route of the drug are available (tablets, sachets and vials) which are all bioequivalent [55].

Pidotimod has a good safety profile without increased frequency of reported side effects or autoimmune disorders in the pidotimod treated patients. Moreover, a study conducted on mice showed that pidotimod has no mutagenic effects either [56]. However, there is a single report in literature of Henoch-Schönlein Purpura associated with pidotimod therapy [41].

#### Pidotimod in the treatment of airway diseases

Acute respiratory infections (ARIs) are still pandemics despite the introduction of new antibiotics and vaccines which contribute to reduce the risk of mortality and morbidity and they remain widespread and affect both young and elder people [1]. Furthermore, the socio-economic burden of ARIs remains high, considering the cost of symptomatic drugs, antibiotics, hospitalization and the indirect cost of absence from work or loss of school days [57].

#### Viral infections

Most common causes of respiratory tract infections have viral origin especially human rhinoviruses (HRV), adenovirus, parainfluenza virus, respiratory syncytial virus (RSV), enterovirus, human metapneumovirus and coronavirus in addition to influenza viruses. [58, 59]. Viruses tend to cause direct invasion of the epithelial cell in the respiratory tract, the most common example of that is HRV which uses intercellular adhesion molecule (ICAM)-1 to gain access to human cells [60], with subsequent stimulation of production of pro-inflammatory cytokines including IL8, IL6, MCP-1 (a chemokine that recruits monocytes) leading to generation of inflammatory response, necrosis of host infected cells [61, 62], decreased ciliary clearance and increase mucous secretions. All these events will eventually cause subsequent obstruction of respiratory tract and impending drainage of secretions [63]. Impaired mucociliary escalator is a good media for bacteria proliferation and secondary infection (Particularly *streptococcus pneumoniae*, *Haemophilus influenzae*, and *Staphylococcus aureus*), causing increase in disease severity, mortality and morbidity with progression to unfavoured *sequela* such as pneumonia [64].

On the other hand, the host immune system tends to control viral infections via induction of apoptosis of infected cells using different mechanisms including: increasing TNF-alpha secretion, stimulation of natural killer (NK) cells to secrete perforin (making pores in the infected cells resulting in induction of cellular apoptosis) and stimulation of macrophages and neutrophils to produce reactive oxygen species causing oxidation of the infected host cell's proteins, lipids and DNA leading to its death [65–67].

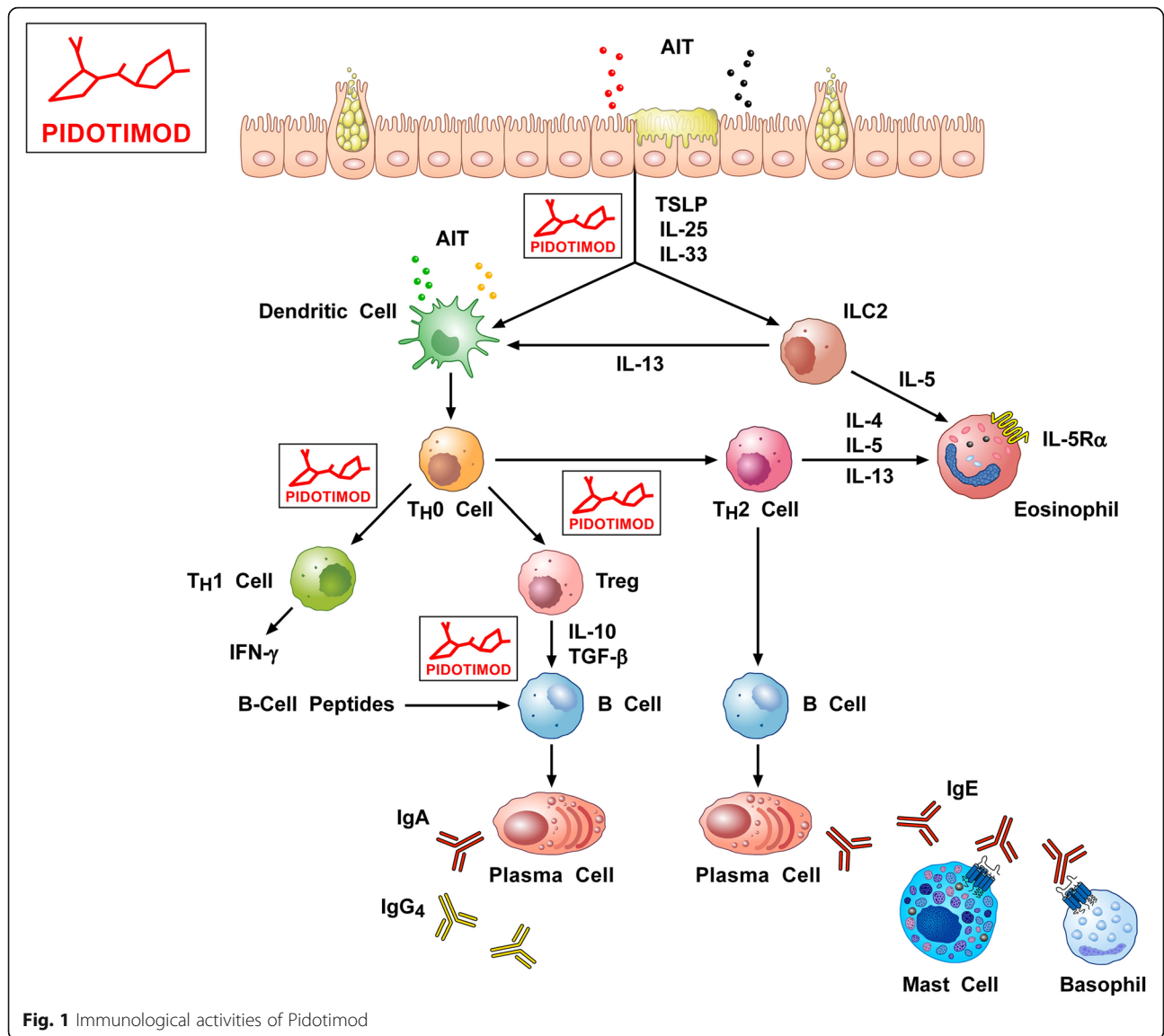
Carta et al., held a study to assess the ability of pidotimod to induce cellular changes enhancing the host immune system to defend infections. This study has documented that pidotimod is able to upregulate TLR2, with no increase in ICAM or IL8 levels, thus playing a protective role in decreasing susceptibility to HRV infection and from neutrophil-mediated damage to the airway surface [48].

Another *in vitro* study aiming to evaluate the effect of pidotimod on immune system demonstrated that it is able to down-regulate MCP-1, which is a master regulator in inflammatory response associated with severe recurrent viral bronchiolitis in the children [68]. Moreover, it is able to upregulate NLRP12, which is a protective molecule against viral-induced abnormal inflammatory response [69].

#### Lower respiratory tract infections

Lower respiratory tract infections (LRTIs), such as acute bronchitis and pneumonia, represent the most common cause of death from infectious diseases and the fourth





**Fig. 1** Immunological activities of Pidotimod

overall most common cause of death worldwide. In 2016, it has been estimated that LRTIs have caused 2 million and 370 thousands deaths, among which more than 1 million were due to *S. pneumoniae* infection. Even if the incidence of LRTIs has decreased in the last decade in the pediatric population, LRTIs remain a leading, albeit preventable, cause of death among elderly patients [57].

Two different studies were planned to evaluate the *in vivo* immunomodulatory effect of pidotimod during acute community-acquired pneumonia (CAP) [70, 71]. One was held on children and the other was held on adults. In both studies, the patients were divided in two groups, one group receiving antibiotics only while the other antibiotics plus Pidotimod. Pidotimod induced the enforcement of the immune system stimulating some proteins such as lactoferrin, cathepsin G and

myeloperoxidase, known to be endowed with potent antibacterial; similarly important is the finding that Pidotimod was associated with a reduced production of TNF-alpha, a proinflammatory cytokine whose excessive production is known as a negative prognostic factor in CAP. Finally, the finding that Pidotimod increased the expression of CD80 and CD86 on DC, confirms its role in triggering the adaptive immunity response [71]. Both studies indicated that pidotimod significantly increased the natural immune system response to an infectious stimulus via stimulation of DC maturation and increased TNF $\alpha$  and IL12 secretion. Pidotimod was found also to determine a long-term enhancement of the immune system activity, up-regulating the expression of CCL3, CXCL1, CXCL2, IL-18, IL-1b, IL-6, IL-8, NFkB1, and NLRP3 genes involved in inflammation and chemotaxis, beside genes

involved in antimicrobial activity (e.g., cAMP, lactoferrin, cathepsin G and Myeloperoxidase), thus reducing the risk of early recurrences during CAP. [70, 71] These were pilot projects on small number of patients and with immunological (and not clinical) primary endpoints.

A recently published meta-analysis assessed a total of 29 RCTs consisting of 4,344 paediatric patients. Ten RCTs were published from Italy, Russia or Greece, and 19 RCTs were published by Chinese groups. Nonetheless, appropriate randomization methods were only used in 15 trials. Only one study had explicit allocation concealment. Since only eight RCTs were double-blind and placebo controlled, the evidence was not assessed as high quality. The meta-analysis indicates that treatment with Pidotimod resulted in a significant increase in the proportion of participants who had lower respiratory tract infections (RR 1.59; 95% CI 1.45–1.74,  $p < 0.00001$ ) compared with the conventional treatment. Pidotimod could significantly decrease the duration of cough and fever. The number of patients using antibiotics was also remarkably decreased in the Pidotimod treatment group. Moreover, Pidotimod administration improved the levels of serum immunoglobulin (IgG, IgA, or IgM) and T-lymphocyte subtypes (CD3+, CD4+). Besides, Pidotimod administration did not increase the risk of adverse events of any cause (RR = 1.05, 95% CI 0.72–1.54,  $p = 0.80$ ) [72].

### Chronic obstructive pulmonary disease (COPD)

COPD is a chronic inflammatory airway disease characterized by chronic fixed airway obstruction [73]. It affects more than 5 % of the population and is associated with high morbidity and mortality [74]. It is the third-ranked cause of death in the United States, causing more than 120,000 deaths annually [75], and it is one of the main causes of morbidity, clinician office visits, hospital admission and loss of working day in industrialized countries [76].

COPD results from complex interactions between environmental (mainly tobacco smoking and/or other pollutants) and molecular risk factors. Molecular risk factors for COPD include a plethora of gene polymorphisms, dysregulations of the antioxidants pool, metalloproteinase abnormalities and uncontrolled role of elastase. COPD exacerbations are triggered most often by respiratory viral infections, mainly rhinoviruses, even if bacterial component is present [77–79]. Moreover, exacerbations tend to be more severe and prolonged if caused by viral pathogens [80].

In terms of immunological response, aside from the inflammation, there is usually a neutrophilic recruitment; sputum eosinophilia can be also relevant, in particular during viral infections [81].

Being the reduction of exacerbation frequency and intensity paramount for the disease control, it appears fundamental to act on the immune system in order to prevent infections and to reach the optimum of care.

Due to its role of potentiating and regulating both the innate and acquired immune systems, Pidotimod appears to be promising also in COPD. In two Italian double-blind randomized trials Pidotimod showed significant reduction in infective COPD exacerbations in Pidotimod treated patients [82, 83]. This positive effect was not only present during the treatment period but also during the follow up, suggesting a protecting action of the drug and a possible use in cyclic regimen [83]. The long-lasting effect of Pidotimod was furtherly confirmed as it demonstrated to potentiate the immune response up to 5 weeks after infection [84].

Pidotimod was found effective also in a particularly susceptible population: elderly COPD patients; a population patients with severe COPD with different degrees of immunological signs of “immunosenescence” achieved a reduced number of exacerbations by the concomitant use of Pidotimod with flu vaccination compared to those treated with vaccine only [85].

In terms of bacterial exacerbations, another study showed that patients treated with Pidotimod plus amoxicillin/ clavulanic acid, had a faster remission of symptoms compared to those treated with antibiotics only [86]. Moreover, Pidotimod was seen able to increase the production of secretory IgA in patients with COPD, contributing in defining Pidotimod as a potential strategy to prevent infections [87].

### Asthma

Asthma phenotypes and endotypes based on inflammatory airway involvement have nowadays increasing medical interests thanks to the development of targeted therapies that aim to hamper directly the mechanism of the underlying pathologic processes [88]. At least two big groups of endotypes have been so far identified: an endotype characterized by Th2 cytokines overexpression (particularly IL5, IL4 and IL13) with concomitant airway eosinophilic inflammation, and a Th2-low inflammatory endotype in which neutrophils are the predominant inflammatory cells within the airways [89]. Th2-high endotypes of asthma can be further distinguished in allergic and non allergic but eosinophilic asthma [90]. The former is the result of an allergic reaction and more typically has a childhood onset, while the latter is the result of an eosinophilic inflammation toward an unknown trigger, it has typically an adult onset and is often associated to chronic rhinosinusitis with nasal polyposis and aspirin hypersensitivity [90]. All the above mentioned endotypes can present in a wide range of severity, and about 5–10% of all asthmatics are classified as affected

by severe asthma as they require high dose inhaled corticosteroids (ICS) plus a second controller and/or systemic glucocorticoids to prevent asthma from becoming 'uncontrolled' or which remains 'uncontrolled' despite this therapy [91]. One of the peculiar feature of severe asthma is the high frequency of exacerbations, often associated with respiratory tract infections, and accounting for the major fraction of the total health-care related costs of asthma. [92] Moreover, a personalized approach to asthma cannot prescind from the diagnosis and treatment of comorbidities, in particular rhinitis and chronic rhinosinusitis with (CRSwNP) or without nasal polyps (CRSsNP). Recently, a cross-sectional study showed that CRS-related exacerbations is associated to lost productivity in asthmatics assessed as number of lost days of work or school [93]. Another common comorbidity of severe asthma is bronchiectasis [94] that it is estimated to be present in up to 25% of patients [95–97]; bronchiectasis is as an irreversible enlargement of the airways, which brings about a predisposition to develop recurrent and often severe lower respiratory tract infections, triggering asthma exacerbations [94]. Into this complex context in which patients with asthma, particularly if severe, are clearly more prone to develop exacerbations linked to upper and/or lower airway infections, an immunostimulatory approach with Pidotimod may be beneficial.

Pre-clinical evidence are already available: in a study investigating the ability of Pidotimod to affect in vitro the phenotype and cytokine profile of blood cells in relation to atopic asthma showed that it was able to down-regulate the expression of CD30 on mononuclear cells isolated from both atopic asthmatic children and healthy controls [51]. Because CD30 has been associated with Th-2 cells, this observation supports the possibility of Pidotimod being able to affect the Th-1/Th-2 balance in atopic asthma.

T2-low asthma endotype patients tend to not react to targeted therapies for pathologic type 2 inflammation, such as glucocorticosteroids. Some, recent studies support the theory that NK cell cytotoxic function is disabled in part by steroids in severe asthma. These NK cells express specialized pro-resolving mediators (SPMs) that are pivotal signals for the resolution of tissue inflammation; nevertheless, SPM abundance or signalling receptors are disrupted in chronic inflammatory disease. SPM receptors and SPMs can in some instances counter the deleterious effects of corticosteroids on the effector function of NK cells. Together, these findings suggest that some severe asthma, patients already refractory to the beneficial actions of corticosteroids, may be harmed by the steroids, increasing susceptibility to viral infection and asthma exacerbations. Thus, leaving an unveiled indication for the use of Pidotimod in these patients once its acts in the prevention of viral infections and asthma exacerbations [98].

## Allergy

In Europe, approximately 23% of the population is affected by allergic rhinitis (AR) [99] that is frequently accompanied by allergic asthma (AA), the prevalence of which increases from less than 2% in individuals without AR to 10–40% in those with AR; this combination of upper and lower respiratory symptoms increases the overall impact on the patient. There is now considerable evidence that the symptoms of AR and AA negatively affect patients' health related quality of life [100–102].

Allergen immunotherapy (AIT) is nowadays the only treatment specifically targeting the allergic inflammatory pathways and it represents a prototype of personalized medicine approach to respiratory disease patients [88]. AIT is highly effective and may induce long-term remission of symptoms. However, ongoing problems include the duration of treatment, the safety of immunotherapy, its cost and accessibility [103].

Adjuvants have the potential to modify the pharmacological and immunological effects of allergen vaccines. They may modulate allergen delivery, act as a depot, stimulate immune responses or limit antibody responses in order to reduce unwanted side effects.

Adjuvants may be used in combination with potentially cumulative effects. Therefore, since Pidotimod action pathways include an effect against TSLP and TNF- $\alpha$  action (Fig. 1), it could open an interesting and unexplored option of adjuvant treatment in allergen immunotherapy depending on the choose route of administration.

## Potential use of Pidotimod as adjuvant in vaccination

A recent study, used Pidotimod in chickens vaccinated with Newcastle disease virus vaccine revealing that Pidotimod could significantly promote growth performance, lymphocyte proliferation, enhance serum antibody titer, CD4/CD8 cell ratios and improve serum IL-2 and IFN- $\gamma$  concentrations, indicating that it significantly improves the immune efficacy of Newcastle disease vaccine, once again reinforcing the role of immunostimulant that already is well established in previous literature findings [104].

## Conclusions

In the last years several immunostimulants of natural or synthetic origins, and working with different mechanisms, have been created for the prevention of RRTIs [1]. Clinical experiences show that there is good evidence about the role of immunostimulants as adjuvant treatment of respiratory diseases in children, while far less exists for adults. Among the currently available immunostimulants, Pidotimod have the potential characteristics to

be one of the most effective adjuvant strategies for respiratory diseases.

Further high-quality, large-scale randomized controlled studies and real-life experiences are essential to provide confirmatory evidence.

#### Abbreviations

AA: Allergic asthma; AIT: Allergen immunotherapy; AR: Allergic rhinitis; ARIs: Acute respiratory infections; BRMs: Biological Response Modifiers; CAP: Community-acquired pneumonia; COPD: Chronic obstructive pulmonary disease; CRSsNP: Chronic rhinosinusitis without nasal polyps; CRSwNP: Chronic rhinosinusitis with nasal polyps; DC: Dendritic cells; HRV: Human rhinoviruses; ICAM: Intercellular adhesion molecule; IL2R: Interleukin-2 receptor; LPS: Lipopolysaccharide; LRTIs: Lower respiratory tract infections; MPLA: Monophosphoryl lipid A; NK: Natural killer; NTHi: Nontypeable *Haemophilus influenzae*; PAMPs: Pathogen-associated molecular patterns; PCML: Polyvalent chemical bacterial lysates; PMBL: Polyvalent mechanical bacterial lysates; RRTI: Recurrent respiratory tract infections; SNS: Supranuclear stria; SPM: Specialized pro-resolving mediators; TLR: Toll like Receptor; URTI: Upper respiratory tract infection

#### Acknowledgements

Not applicable.

#### Authors' contributions

All the Authors contributed in revising the literature and in writing the manuscript. All authors read and approved the final manuscript.

#### Funding

No funds related with this article.

#### Availability of data and materials

Not applicable.

#### Ethics approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interests. GWC is member of the Editorial Board of *Multidisciplinary Respiratory Medicine*.

#### Author details

<sup>1</sup>Department of Biomedical Sciences, Humanitas University, Pieve Emanuele, MI, Italy. <sup>2</sup>Personalized Medicine, Allergy and Asthma - Humanitas Clinical and Research Center - IRCCS, Via Alessandro Manzoni 56, 20089 Rozzano, MI, Italy. <sup>3</sup>Central Hospital of Funchal, SESARAM, EPE, Madeira, Portugal. <sup>4</sup>Ain Shams University, Faculty of Medicine, Cairo, Egypt. <sup>5</sup>Department of Translational Medical Sciences and Center for Basic and Clinical Immunology Research, University of Naples Federico II, Naples, Italy.

Received: 24 May 2019 Accepted: 23 July 2019

Published online: 04 November 2019

#### References

- Ferrario BE, Garuti S, Braido F, Canonica GW. Pidotimod: the state of art. *Clin Mol Allergy*. 2015;13:8.
- Lanzilli G, Falchetti R, Tricarico M, Ungheri D, Fuggetta MP. In vitro effects of an immunostimulating bacterial lysate on human lymphocyte function. *Int J Immunopathol Pharmacol*. 2005;18:245–54.
- Morandi B, Agazzi A, D'Agostino A, Antonini F, Costa G, Sabatini F, et al. A mixture of bacterial mechanical lysates is more efficient than single strain lysate and of bacterial-derived soluble products for the induction of an activating phenotype in human dendritic cells. *Immunol Lett*. 2011;138:86–91.
- Huber M, Mossmann H, Bessler WG. Th1-orientated immunological properties of the bacterial extract OM-85-BV. *Eur J Med Res*. 2005;10:209–17.
- Del-Rio-Navarro BE, Espinosa Rosales F, Flenady V, Sienra-Monge JJ. Immunostimulants for preventing respiratory tract infection in children. *Cochrane Database Syst Rev*. 2006;4:CD004974.
- Schaad UB. OM-85 BV, an immunostimulant in pediatric recurrent respiratory tract infections: a systematic review. *World J Pediatr*. 2010;6:5–12.
- Gutiérrez-Tarango MD, Berber A. Safety and efficacy of two courses of OM-85 BV in the prevention of respiratory tract infections in children during 12 months. *Chest*. 2001;119:1742–8.
- Gómez Barreto D, De la Torre C, Alvarez A, Faure A, Berber A. Safety and efficacy of OM-85-BV plus amoxicillin/clavulanate in the treatment of subacutesinusitis and the prevention of recurrent infections in children. *Allergol Immunopathol (Madr)*. 1998;26:17–22.
- Macchi A, Vecchia LD. Open comparative, randomized controlled clinical study of a new immunostimulating bacterial lysate in the prophylaxis of upper respiratory tract infections. *Arzneimittelforschung*. 2005;55:276–81.
- Braido F, Melioli G, Cazzola M, Fabbri L, Blasi F, Moretta L, et al. Sub-lingual administration of a polyvalent mechanical bacterial lysate (PMBL) in patients with moderate, severe, or very severe chronic obstructive pulmonary disease (COPD) according to the GOLD spirometric classification: a multicentre, double-blind, randomised, controlled, phase IV study (AIACE study: advanced immunological approach in COPD exacerbation). *Pulm Pharmacol Ther*. 2015;33:75–80.
- De la Rosa G, Yang D, Tewary P, Varadhachary A, Oppenheim JJ. Lactoferrin acts as an Alarmin to promote the recruitment and activation of APCs and antigen-specific immune responses. *J Immunol*. 2008;180:6868–76.
- Puddu P, Latorre D, Carollo M, Catizone A, Ricci G, Valenti P, et al. Bovine Lactoferrin counteracts toll-like receptor mediated activation signals in antigen presenting cells. *PLoS One*. 2011;6:e22504.
- Bournazou I, Mackenzie KJ, Duffin R, Rossi AG, Gregory CD. Inhibition of eosinophil migration by lactoferrin. *Immunol Cell Biol*. 2010;88:220–3.
- He S, McEuen AR, Blewett SA, Li P, Buckley MG, Leufkens P, et al. The inhibition of mast cell activation by neutrophil lactoferrin: uptake by mast cells and interaction with tryptase, chymase and cathepsin G. *Biochem Pharmacol*. 2003;15(65):1007–15.
- Fischer R, Debbabi H, Dubarry M, Boyaka P, Tomé D. Regulation of physiological and pathological Th1 and Th2 responses by lactoferrin. *Biochem Cell Biol*. 2006;84:303–11.
- Manzoni P, Rinaldi M, Cattani S, Pugni L, Romeo MG, Messner H, et al. Bovine lactoferrin supplementation for prevention of late-onset sepsis in very low-birth-weight neonates: a randomized trial. *JAMA*. 2009;302:1421–8.
- Euba B, López-López N, Rodríguez-Arce I, Fernández-Calvet A, Barberán M, Caturla N, et al. Resveratrol therapeutics combines both antimicrobial and immunomodulatory properties against respiratory infection by nontypeable *Haemophilus influenzae*. *Sci Rep*. 2017;16:7–12860.
- Su Q, Pu H, Hu C. Neuroprotection by combination of resveratrol and enriched environment against ischemic brain injury in rats. *Neuro Res*. 2016;38:60–8.
- Lanzilli G, Cottarelli A, Nicotera G, Guida S, Ravagnan G, Fuggetta MP. Anti-inflammatory effect of resveratrol and polydatin by in vitro IL-17 modulation. *Inflammation*. 2012;35:240–8.
- Gertz M, Nguyen GT, Fischer F, Suenkel B, Schlicker C, Fränzel B, et al. A molecular mechanism for direct Sirtuin activation by resveratrol. *PLoS One*. 2012;7:e49761.
- Yao J, Wei C, Wang JY, Zhang R, Li YX, Wang LS. Effect of resveratrol on Treg/Th17 signaling and ulcerative colitis treatment in mice. *World J Gastroenterol*. 2015;21:6572–81.
- Knobloch J, Wahl C, Feldmann M, Jungck D, Strauch J, Stoelben E, et al. Resveratrol attenuates the release of inflammatory cytokines from human bronchial smooth muscle cells exposed to Lipoteichoic acid in chronic obstructive pulmonary disease. *Basic Clin Pharmacol Toxicol*. 2014;114:202–9.
- Andrews CS, Matsuyama S, Lee BC, Li JD. Resveratrol suppresses NTHi-induced inflammation via up-regulation of the negative regulator MyD88 short. *Sci Rep*. 2016;6:34445.
- Wu S, Huang J. Resveratrol alleviates *Staphylococcus aureus* pneumonia by inhibition of the NLRP3. *Exp Ther Med*. 2017;14:6099–104.
- De Becker G, Moulin V, Pajak B, Bruck C, Francotte M, Thiriart C, et al. The adjuvant monophosphoryl lipid A increases the function of antigen-presenting cells. *Int Immunol*. 2000;12:807–15.
- Hepner DG, Gordon DM, Gross M, Wellde B, Leitner W, Krzych U, et al. Safety, immunogenicity, and efficacy of plasmodium falciparum repeatless



- circumsporozoite protein vaccine encapsulated in liposomes. *J Infect Dis*. 1996;174:361–6.
27. Thoelen S, Van Damme P, Mathei C, Leroux-Roels G, Desombere I, Safary A, et al. Safety and immunogenicity of a hepatitis B vaccine formulated with a novel adjuvant system. *Vaccine*. 1998;16:708–14.
  28. Konno R, Dobbelaere KO, Godeaux OO, Tamura S, Yoshikawa H. Immunogenicity, reactogenicity, and safety of human papillomavirus 16/18 AS04-adjuvanted vaccine in Japanese women: interim analysis of a phase II, double-blind, randomized controlled trial at month 7. *Int J Gynecol Cancer*. 2009;19:905–11.
  29. Puggioni F, Durham SR, Francis JN. Monophosphoryl lipid a (MPL) promotes allergen-induced immune deviation in favour of Th1 responses. *Allergy*. 2005;60:678–84.
  30. Rosewich M, Schulze J, Eickmeier O, Telles T, Rose MA, Schubert R, et al. Tolerance induction after specific immunotherapy with pollen allergoids adjuvanted by monophosphoryl lipid a in children. *Clin Exp Immunol*. 2010;160:403–10.
  31. Romero CD, Varma TK, Hobbs JB, Reyes A, Driver B, Sherwood ER. The toll-like receptor 4 agonist Monophosphoryl lipid a augments innate host resistance to systemic bacterial infection. *Infect Immun*. 2011;79:3576–87.
  32. Astiz ME, Galera A, Saha DC, Carpati C, Rackow EC. Monophosphoryl lipid a protects against gram-positive sepsis and tumor necrosis factor. *Shock*. 1994;2:271–4.
  33. Zuccotti GV, Mameli C. Pidotimod: the past and present. *Ital J Pediatr*. 2013;39:75.
  34. Giagulli C, Noerder M, Avolio M, Becker PD, Fiorentini S, Guzman CA, et al. Pidotimod promotes functional maturation of dendritic cells and displays adjuvant properties at the nasal mucosa level. *Int Immunopharmacol*. 2009;9:1366–73.
  35. Trabattoni D, Clerici M, Centanni S, Mantero M, Garziano M, Blasi F. Immunomodulatory effects of pidotimod in adults with community-acquired pneumonia undergoing standard antibiotic therapy. *Pulm Pharmacol Ther*. 2017;44:24–9.
  36. Riboldi P, Gerosa M, Meroni PL. Pidotimod: a reappraisal. *Int J Immunopathol Pharmacol*. 2009;22:255–62.
  37. De la Torre GC, Pacheco Ríos A, Escalante Domínguez AJ, del Río Navarro BE. Comparative meta-analysis of immunoestimulant agents used in pediatric patients in Mexico. *Rev Alerg Mex*. 2005;52:25–38.
  38. Careddu P, Alfano S, Zavattini G. Pidotimod III the prophylaxis of recurrent acute tonsillitis III childhood. *Adv Ther*. 1992;9:174–83.
  39. Marelli P, Lupetti A SS, Uslenghi C, Girardello R, Campa M. Potenzamento della resistenza alle infezioni virali e batteriche dopo somministrazione di pidotimod nel topo. *Drugs Exp Clin Res*. 1993;XIX:15–21.
  40. Santamaria F, Montella S, Stoccheri M, Pirillo P, Bozzetto S, Giordano G, Poeta M, Baraldi E. Effects of pidotimod and bifidobacteria mixture on clinical symptoms and urinary metabolomic profile of children with recurrent respiratory infections: A randomized placebo-controlled trial. *Pulm Pharmacol Ther*. 2019;58:101818.
  41. Cantarini L, Brogna A, Fioravanti A, Galeazzi M. Henoch-Schönlein purpura associated with pidotimod therapy. *Clin Exp Rheumatol*. 2008;26:S152.
  42. Burgio GR, Marseglia GL, Severi F, De Benedetti F, Masarone M, Ottolenghi A, et al. Immunoactivation by pidotimod in children with recurrent respiratory infections. *Arzneimittelforschung*. 1994;44(12A):1525–9.
  43. Careddu P. Role of immunoactivation with pidotimod in recurrent respiratory infections in childhood. *Arzneimittelforschung*. 1994;44(12A):1506–11.
  44. Akira S, Takeda K, Kaisho T. Toll-like receptors: critical proteins linking innate and acquired immunity. *Nat Immunol*. 2001;2:675–80.
  45. Akira S, Takeda K. Toll-like receptor signalling. *Nat Rev Immunol*. 2004;4:499–511.
  46. Gilmore TD. Introduction to NF- $\kappa$ B: players, pathways, perspectives. *Oncogene*. 2006;25:6680–4.
  47. Brasier AR. The NF- $\kappa$ B regulatory network. *Cardiovasc Toxicol*. 2006;6:111–30.
  48. Carta S, Silvestri M, Rossi GA. Modulation of airway epithelial cell functions by Pidotimod: NF- $\kappa$ B cytoplasmic expression and its nuclear translocation are associated with an increased TLR-2 expression. *Ital J Pediatr*. 2013;39:29.
  49. Pugliese A, Uslenghi C, Torre D, D'Alonzo L, Corgnati F, Girardello R. Evaluation of the effect of pidotimod on the in vitro production of interferons. *Int J Immunother*. 1995;11:71–6.
  50. Zhou Y, Dai Y. Comparison of effects of Pidotimod and spleen aminopeptide on clinical symptoms and Th1/Th2 cytokines in children with RRI. *Chin J Biochem Pharm*. 2012;33:64–9.
  51. Gourgiotis D, Papadopoulos NG, Bossios A, Zamanis P, Saxoni-Papageorgiou P. Immune modulator pidotimod decreases the in vitro expression of CD30 in peripheral blood mononuclear cells of atopic asthmatic and normal children. *J Asthma*. 2004;41:285–7.
  52. Zuccotti GV, Mameli C, Trabattoni D, Beretta S, Biasin M, Guazzarotti L, et al. Immunomodulating activity of Pidotimod in children with Down syndrome. *J Biol Regul Homeost Agents*. 2013;27:253–8.
  53. Lokshina EE, Kravchenko OV, Zaytseva OV. Pidotimod in treatment of children with acute respiratory infection with concomitant recurrent obstructive syndrome. *Curr Paediatr*. 2011;10:34–41.
  54. Macchi A, Terranova P, Digilio E, Simoncini D, Peirola A, Castelnovo G. Nasal cell vitality: the Pidotimod activity in children's recurrent respiratory infections. *Rhinologist*. 2017;3:15–23.
  55. Mailland F, Coppi G, Silingardi S. Pharmacokinetics and oral bioavailability of pidotimod in humans. *Arzneimittelforschung*. 1994;44(12A):1465–9.
  56. Adams K, Allen JA, Brooker PC, Henderson L, Jones E, Proudlock RJ, et al. Genotoxicity testing of Pidotimod in vivo and in vitro. *Arzneim Forsch Drug Res*. 1994;44 II:1454–9.
  57. GBD 2016 LRI Collaborators. Estimates of the global, regional, and national morbidity, mortality, and aetiologies of lower respiratory infections in 195 countries, 1990–2016: a systematic analysis for the global burden of disease study 2016. *Lancet Infect Dis*. 2018;18:30310–4.
  58. Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, Aboyans V, et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the global burden of disease study 2010. *Lancet*. 2012;380:2095–128.
  59. Heikkinen T, Järvinen A. The common cold. *Lancet*. 2003;361:51–9.
  60. Othumpangat S, Regier M, Piedimonte G. Nerve growth factor modulates human rhinovirus infection in airway epithelial cells by controlling ICAM-1 expression. *Am J Physiol Lung Cell Mol Physiol*. 2012;302:1057–66.
  61. De Jong MD, Simmons CP, Thanh TT, Hien VM, Smith GJ, Chau TN, et al. Fatal outcome of human influenza A (H5N1) is associated with high viral load and hypercytokinemia. *Nat Med*. 2006;12:1203–7.
  62. Kobasa D, Jones SM, Shinya K, Kash JC, Copps J, Ebihara H, et al. Aberrant innate immune response in lethal infection of macaques with the 1918 influenza virus. *Nature*. 2007;445:319–23.
  63. Dasaraju PV, Liu C. Chapter 93: infections of the respiratory system. In: Baron S, editor. *Medical Microbiology*. 4th ed. Galveston: University of Texas Medical Branch at Galveston; 1996.
  64. Smith AM, McCullers JA. Secondary bacterial infections in influenza virus infection pathogenesis. *Curr Top Microbiol Immunol*. 2014;385:327–56.
  65. Topham NJ, Hewitt EW. Natural killer cell cytotoxicity: how do they pull the trigger? *Immunology*. 2009;128:7–15.
  66. Kash JC, Xiao Y, Davis AS, Walters KA, Chertow DS, Easterbrook JD, et al. Treatment with the reactive oxygen species scavenger EUK-207 reduces lung damage and increases survival during 1918 influenza virus infection in mice. *Free Radic Biol Med*. 2013;67:235–47.
  67. Kash JC, Taubenberger JK. The role of viral, host, and secondary bacterial factors in influenza pathogenesis. *Am J Pathol*. 2015;185:1528–36.
  68. Welliver RC, Garofalo RP, Ogra PL. Beta-chemokines, but neither T helper type 1 nor T helper type 2 cytokines, correlate with severity of illness during respiratory syncytial virus infection. *Pediatr Infect Dis J*. 2002;21:457–61.
  69. Fogli M, Caccuri F, Iaria ML, Giagulli C, Corbellini S, Campilongo F, et al. The immunomodulatory molecule pidotimod induces the expression of the NOD-like receptor NLRP12 and attenuates TLR-induced inflammation. *J Biol Regul Homeost Agents*. 2014;28:753–66.
  70. Esposito S, Garziano M, Rainone V, Trabattoni D, Biasin M, Senatore L, et al. Immunomodulatory activity of pidotimod administered with standard antibiotic therapy in children hospitalized for community-acquired pneumonia. *J Transl Med*. 2015;13:288.
  71. Trabattoni D, Clerici M, Centanni S, Mantero M, Garziano M, Blasi F. Immunomodulatory effects of pidotimod in adults with community-acquired pneumonia undergoing standard antibiotic therapy. *Pulm Pharmacol Ther*. 2017;44:24–9.
  72. Niu H, Wang R, Jia YT, Cai Y. Pidotimod, an immunostimulant in pediatric recurrent respiratory tract infections: a meta-analysis of randomized controlled trials. *Int Immunopharmacol*. 2019;67:35–45.
  73. Wedzicha J. Mechanisms of chronic obstructive pulmonary disease exacerbations. *Ann Am Thorac Soc*. 2015;12:S157–9.

74. Global Initiative for Chronic Obstructive Lung Disease (GOLD) - <https://goldcopd.org>. Accessed the 16 July 2019.
75. Miniño A, Murphy S, Xu J, Kochanek K. Deaths: final data for 2008. *Natl Vital Stat Rep*. 2018;59(10):1–126 Available at: [https://www.cdc.gov/nchs/data/nvsr/nvsr59/nvsr59\\_10.pdf](https://www.cdc.gov/nchs/data/nvsr/nvsr59/nvsr59_10.pdf). Accessed 17 Nov 2018.
76. Buist A, McBurnie M, Vollmer W, Gillespie S, Burney P, Mannino D, et al. International variation in the prevalence of COPD (the BOLD study): a population-based prevalence study. *Lancet*. 2007;370:741–50.
77. Seemungal TA, Donaldson GC, Paul EA, Bestall JC, Jeffries DJ, Wedzicha JA. Effect of exacerbation on quality of life in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 1998;157:1418–22.
78. Wedzicha JA, Seemungal TA. COPD exacerbations: defining their cause and prevention. *Lancet*. 2007;370:786–96.
79. George SN, Garcha DS, Mackay AJ, Patel AR, Singh R, Sapsford RJ, et al. Human rhinovirus infection during naturally occurring COPD exacerbations. *Eur Respir J*. 2014;44:87–96.
80. Aaron SD, Donaldson GC, Whitmore GA, Hurst JR, Ramsay T, Wedzicha JA. Time course and pattern of COPD exacerbation onset. *Thorax*. 2012;67:238–43.
81. Papi A, Bellettato CM, Braccioni F, Romagnoli M, Casolari P, Caramori G, et al. Infections and airway inflammation in chronic obstructive pulmonary disease severe exacerbations. *Am J Respir Crit Care Med*. 2006;173:1114–21.
82. Bisetti G, Ciappi G, Bariffi F, Catena E, Rocco V, Vaccaro L, et al. Evaluation of the efficacy of Pidotimod on the exacerbations in patients affected with chronic bronchitis. *Arzneim Forsch Drug Res*. 1994;44 II:1499–502.
83. Ciaccia A, et al. Pidotimod activity against chronic bronchitis exacerbations. *Arzneim Forsch Drug Res*. 1994;44:1516–20.
84. Benetti GP, Illeni MT, Passera A, Bombelli G, Lavecchia G, Uslenghi C. Ex vivo evaluation of Pidotimod activity in patients with chronic obstructive. *Arzneimittelforschung*. 1994;44:1503–5.
85. Chen J, Wang H, Liu Y, Xia C, Qi M. Immunological function of T-lymphocyte in the elderly with chronic obstructive pulmonary disease during acute exacerbations medication intervention. *Chin J Geriatr*. 2006;25:434–40.
86. Pozzi E, Dolcetti A, Orlandi O, Cirianni C, Moreo G, Piacenza G, et al. Pidotimod in the treatment in patients affected by bacterial exacerbations of chronic bronchitis. *Arzneim Forsch Drug Res*. 1994;44:1495–8.
87. Cogo R, Ramponi A, Monzani G, Uslenghi C, Girardello R. Valutazione dell'attività del Pidotimod sulla secrezione di IgA in pazienti affetti da BPCO. *Giorn It Allergol Immunol Clin*. 1993;3:253–9.
88. Canonica GW, Ferrando M, Baiardini I, Puggioni F, Racca F, Passalacqua G, et al. Asthma: personalized and precision medicine. *Curr Opin Allergy Clin Immunol*. 2018;18:51–8.
89. Lambrecht BN, Hammad H. The immunology of asthma. *Nat Immunol*. 2015;16:45–56.
90. Desai M, Oppenheimer J. Elucidating asthma phenotypes and endotypes: progress towards personalized medicine. *Ann Allergy Asthma Immunol*. 2016;116:394–401.
91. Chung KF, Wenzel SE, Brozek JL, Bush A, Castro M, Sterk PJ, et al. International ERS/ATS guidelines on definition, evaluation and treatment of severe asthma. *Eur Respir J*. 2014;43:343–73.
92. Shemesh E, Kleinman LC. Asthma: the past, future, environment, and costs. *J Allergy Clin Immunol*. 2017;140:688–9.
93. Phillips KM, Bergmark RW, Hoehle LP, Caradonna DS, Gray ST, Sedaghat AR. Chronic rhinosinusitis exacerbations are differentially associated with lost productivity based on asthma status. *Rhinology*. 2018;1(56):323–9.
94. Crimi C, Ferri S, Crimi N. Bronchiectasis and asthma: a dangerous liaison? *Curr Opin Allergy Clin Immunol*. 2019;19:46–52.
95. Bisaccioni C, Aun MV, Cajuela E, Kalil J, Agondi RC, Giavina-Bianchi P. Comorbidities in severe asthma: frequency of rhinitis, nasal polyposis, gastroesophageal reflux disease, vocal cord dysfunction and bronchiectasis. *Clinics*. 2009;64:769–73.
96. Porsbjerg C, Menzies-Gow A. Co-morbidities in severe asthma: clinical impact and management. *Respirology*. 2017;22:651–61.
97. Heffler E, Blasi F, Latorre M, Menzella F, Paggiaro P, Pelaia G, et al. The severe asthma network in Italy: findings and perspectives. *J Allergy Clin Immunol Pract*. 2018;S2213-2198(18):30673–1.
98. Duvall MG, Krishnamoorthy N, Levy BD. Non-type 2 inflammation in severe asthma is propelled by neutrophil cytoplasts and maintained by defective resolution. *Allergol Int*. 2018. <https://doi.org/10.1016/j.alit.2018.11.006>.
99. Linneberg A, Dam Petersen K, Hahn-Pedersen J, Hammerby E, Serup-Hansen N, Boxall N. Burden of allergic respiratory disease: a systematic review. *Clin Mol Allergy*. 2016;14:12.
100. Spector SL. Overview of comorbid associations of allergic rhinitis. *J Allergy Clin Immunol*. 1997;99:S773–80.
101. Ozdoganoglu T, Songu M. The burden of allergic rhinitis and asthma. *Ther Adv Respir Dis*. 2012;6:11–23.
102. Linneberg A, Henrik Nielsen N, Frolund L, Madsen F, Dirksen A, Jorgensen T, et al. The link between allergic rhinitis and allergic asthma: a prospective population-based study. The Copenhagen allergy study. *Allergy*. 2002;57:1048–52.
103. Gunawardana NC, Durham SR. New approaches to allergen immunotherapy. *Ann Allergy Asthma Immunol*. 2018;121:293–305.
104. Qu S, Dai C, Qiu M, Zhang R, Wang C, Cui L, et al. Effects of pidotimod soluble powder and immune enhancement of Newcastle disease vaccine in chickens. *Immunol Lett*. 2017;187:14–8.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

